

## Quest Rhod-4™ Calcium Reagents and Screen Quest™ Rhod-4 NW Calcium Assay Kits

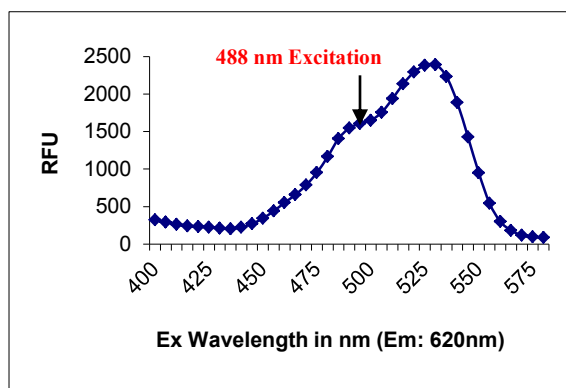
### Introduction

Calcium acts as a universal second messenger in a variety of cells. The beginning of life, the act of fertilization, is regulated by  $\text{Ca}^{2+}$ . Numerous functions of all types of cells are regulated by  $\text{Ca}^{2+}$  to a greater or lesser degree. Since the 1920s, scientists have attempted to measure  $\text{Ca}^{2+}$ , but few were successful due to limited availability of  $\text{Ca}^{2+}$  probes. The first reliable measurements of  $\text{Ca}^{2+}$  were performed by Ridgway and Ashley by injecting the photoprotein aequorin into the giant muscle fiber of the barnacle. Subsequently, in the 1980s, Tsien and colleagues produced a variety of fluorescent indicators. Among them the rhodamine-based  $\text{Ca}^{2+}$  reagents (such as Rhod-2) have been the most valuable longer wavelength dye for measuring  $\text{Ca}^{2+}$  with red fluorescence.

### Quest Rhod-4™ Calcium Indicators, the Most Sensitive Red Fluorescent Calcium Probe

Although Rhod-2 has been the most popular red fluorescent  $\text{Ca}^{2+}$  indicator, its mitochondrial localization and high basal  $\text{Ca}^{2+}$  signal in cells have severely limited its cellular applications. In addition, the less optimal excitation of Rhod-2 at 488 nm makes it less robust to use with some instruments (such as FLIPR™) that have only 488 nm excitation light source. Our Quest Rhod-4™ serial calcium detection reagents have been developed to address these limitations of Rhod-2.

The absorption and emission peaks of Quest Rhod-4™ reagents are 530 nm and 555 nm, respectively. Although Rhod-4 has maximum absorption at 530 nm, its absorption at 488 nm is quite strong (see Figure 1). It is quite unique that Quest Rhod-4™ can be well excited with an argon ion laser at 488 nm besides the longer wavelength excitations at 514 nm, 532 nm and 546 nm. Quest Rhod-4™ emits fluorescence (at 555 nm) which increases with the increasing  $\text{Ca}^{2+}$ . Quest Rhod-4™ is determined to undergo a >200-fold increase in fluorescence upon binding to  $\text{Ca}^{2+}$ . Because the range of increase in  $\text{Ca}^{2+}$  in many cells after stimulation is generally 5- to 10-fold, Quest Rhod-4™ is an excellent probe to use with high sensitivity in this region. The  $K_d$  of Quest Rhod-4™ is estimated to be 525 nM (22 °C, pH 7.0–7.5), but this value may be significantly influenced by pH, viscosity, and binding proteins *in vivo* conditions.



**Figure 1.** Excitation Spectrum of Quest Rhod-4™, sodium salt in the presence of calcium chloride

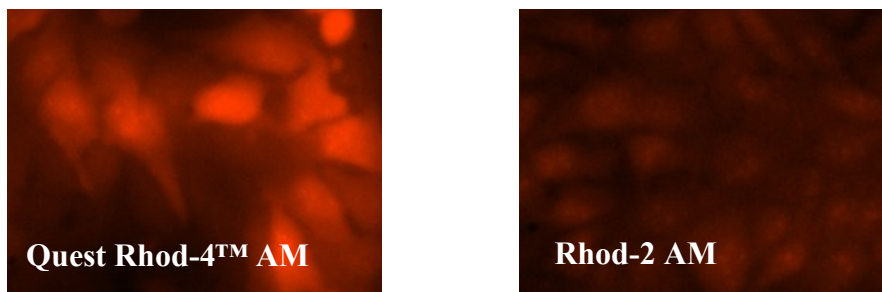
Besides their convenient excitation wavelengths and large fluorescence enhancement by calcium, Quest Rhod-4™ is much brighter in cells than Rhod-2 as shown in Figure 2. More importantly, Quest Rhod-4™ is predominantly localized in cytosols unlike Rhod-2 that is mainly localized in mitochondria. In addition, Quest Rhod-4 AM is much more readily loaded into live cells than Rhod-2 AM. Quest Rhod-4™ reagents have a less temperature-dependent cell loading property, giving similar results either at room temperature or 37 °C. This characteristic makes Quest Rhod-4™ more robust for HTS applications than Rhod-2 AM.

**Table 1.** Spectral and Ca<sup>2+</sup>-Binding Properties of Quest Rhod-4™ Calcium Detection Reagents

Ca <sup>2+</sup> Indicator	Excitation	Emission	K <sub>d</sub> of Ca <sup>2+</sup> -Binding
Quest Rhod-4™	530 nm	555 nm	525 nM

Compared to Rhod-2, our Quest Rhod-4™ calcium detection reagents have the following advantages:

- *Convenient Excitation Wavelengths:* multiple excitation options @ ~490 nm, 514 nm, 532 nm and 546 nm.
- *Much Larger Assay Window:* 10 times larger than Rhod-2 AM.
- *Enhanced Intensity:* 4 times brighter than that of Rhod-2 AM.
- *Flexible Loading:* dye loading at room temperature rather than 37 °C.
- *Versatile Packing Sizes to Meet Your Special Needs:* 1 mg; 10 x 50 µg; 20 x 50 µg; HTS packages.



**Figure 2.** U2OS cells were seeded overnight at 40,000 cells/100 µL/well in a 96-well black wall/clear bottom costar plate. The growth medium was removed, and the cells were incubated with, respectively, 100 µL of Rhod-2 AM (4 µM) and Quest Rhod-4™ AM ((4 µM) in HHBS in a 37 °C, 5% CO<sub>2</sub> incubator for 1 hour. The cells were washed twice with 200 µL HHBS, then imaged with a fluorescence microscope (Olympus IX71) using TRITC channel.

## Use of Quest Rhod-4™ AM Esters

### 1. Load Cell with Quest Rhod-4™ AM Esters:

AM esters are the non-polar esters that readily cross live cell membranes, and rapidly hydrolyzed by cellular esterases inside live cells. AM esters are widely used for loading a variety of polar fluorescent probes into live cell non-invasively. However, cautions must be excised when AM esters are used since they are susceptible to hydrolysis, particularly in solution. They should be reconstituted just before use in high-quality, anhydrous dimethylsulfoxide (DMSO). DMSO stock solutions may be stored desiccated at -20 °C and protected from light. Under these conditions, AM esters should be stable for several months.

Following is our recommended protocol for loading Quest Rhod-4™ AM esters into live cells. This protocol only provides a guideline, and should be modified according to your specific needs.

- Prepare a 2 to 5 mM stock solution of Quest Rhod-4™ AM esters in high-quality, anhydrous DMSO.
- On the day of the experiment, either dissolve Quest Rhod-4™ in DMSO or thaw an aliquot of the indicator stock solution to room temperature. Prepare a working solution of 1 to 10 µM in Hanks and Hepes buffer (HHBS) or the buffer of your choice with 0.02% *Pluronic*® F-127. For most of cell lines, Quest Rhod-4™ reagents with a concentration ranging from 4 to 5 µM are recommended. The exact concentration of the indicator required for cell loading must be determined empirically. To avoid any artifacts caused by overloading and potential dye toxicity, it is recommended to use the minimal probe concentration that can yield sufficient signal strength.

*Note: The nonionic detergent Pluronic® F-127 is sometimes used to increase the aqueous solubility of Quest Rhod-4™ AM esters. A variety of Pluronic® F-127 solutions can be purchased from AAT Bioquest.*

- If your cells containing the organic anion-transporters, probenecid (1–2.5 mM) or sulfipyrazone (0.1–0.25 mM) should be added into the cell medium to reduce leakage of the de-esterified indicators.

*Note: A variety of ReditUse™ probenecid including water soluble sodium salt and stabilized solution can be purchased from AAT Bioquest.*

- d) Incubate cells with the Quest Rhod-4™ AM esters for 30 minutes to one hour at room temperature or 37 °C.  
*Note: Decreasing the loading temperature might reduce the compartmentalization of the indicator.*
- e) Wash cells 1-2 times in HHBS or buffer of your choice (containing an anion transporter inhibitor, such as 2.5 mM probenecid, if applicable) to remove excess probes.
- f) Run the experiments at Ex/Em = 530/570 nm.

## 2. Measure Intracellular Calcium Responses:

To determine either the free calcium concentration of a solution or the  $K_d$  of a single-wavelength calcium indicator, the following equation is used:

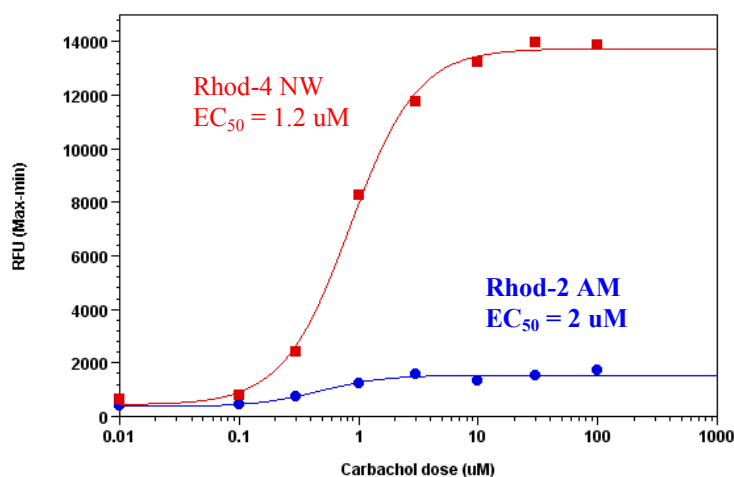
$$[\text{Ca}]_{\text{free}} = K_d[F - F_{\text{min}}]/F_{\text{max}} - F]$$

Where  $F$  is the fluorescence of the indicator at experimental calcium levels,  $F_{\text{min}}$  is the fluorescence in the absence of calcium and  $F_{\text{max}}$  is the fluorescence of the calcium-saturated probe.

The dissociation constant ( $K_d$ ) is a measure of the affinity of the probe for calcium. The Ca-binding and spectroscopic properties of fluorescent indicators vary quite significantly in cellular environments compared to calibration solutions. *In situ* response calibrations of intracellular indicators typically yield  $K_d$  values significantly higher than *in vitro* determinations. *In situ* calibrations are performed by exposing loaded cells to controlled  $\text{Ca}^{2+}$  buffers in the presence of ionophores such as A-23187, 4-bromo A-23187 and ionomycin. Alternatively, cell permeabilization agents such as digitonin or Triton® X-100 can be used to expose the indicator to the controlled  $\text{Ca}^{2+}$  levels of the extracellular medium. The  $K_d$  value of Quest Rhod-4™ is listed in Table 1 for your reference.

## Use of Screen Quest™ Rhod-4 NW Calcium Assay Kits for HTS Applications

GPCR activation can be detected by direct measurement of the receptor mediated cAMP accumulation, or changes in intracellular  $\text{Ca}^{2+}$  concentration. GPCR targets that couple via Gq produce an increase in intracellular  $\text{Ca}^{2+}$  that can be measured by using a combination of Quest Rhod-4™ reagents and a fluorescence microplate reader. The fluorescence imaging plate readers (such as, FLIPR™, FDSS or BMG NovoStar™) have a cooled CCD camera imaging system which collects the signal from each well of a microplate (both 96 and 384-well) simultaneously. These plate readers can read at sub-second intervals, which enables the kinetics of the response to be captured, and has an integrated pipettor that may be programmed for successive liquid additions. Besides their robust applications for GPCR targets, our Screen Quest™ Rhod-4 Calcium Assay Kits can be also used for characterizing calcium ion channels and screening calcium ion channel-targeted compounds.



**Figure 2.** Carbachol Dose Response was measured in HEK-293 cells with Screen Quest™ Rhod-4 NW Assay kit and Rhod-2 AM under the same assay conditions. HEK-293 cells were seeded overnight at 40,000 cells/100  $\mu\text{L}$ /well in a 96-well black wall/clear bottom costar plate. The growth medium was removed, and the cells were incubated with, respectively, 100  $\mu\text{L}$  of the Screen Quest™ Rhod-4 NW calcium assay kit and Rhod-2 AM for 1 hour at room temperature. Carbachol (25  $\mu\text{L}$ /well) was added by NOVOstar™ (BMG LabTech) to achieve the final indicated concentrations. The  $\text{EC}_{50}$  of Rhod-4 NW is about 1.2  $\mu\text{M}$ . The excitation and emission were 530nm and 570nm respectively.

Our Screen Quest™ Rhod-4 Calcium Assay Kits have the following advantages for HTS applications:

- *Longer Wavelengths:* multiple excitations @ 488, 514, 532 & 546 nm; maximum emission @ ~555 nm.
- *No Wash Required and No Quencher Interference with Your Targets.*
- *Robust Performance:* enable calcium assays that are impossible with Rhod-2 AM.
- *Strongest Signal Intensity:* 4 times brighter than that of Rhod-2 AM.
- *Larger Assay Window:* 10 times larger than Rhod-2 AM.

## Conclusions

Because of the importance of  $\text{Ca}^{2+}$  in biology, numerous techniques/methods for analyzing the mechanisms of cellular and/or subcellular  $\text{Ca}^{2+}$  activity have been established. Although each method for analyzing  $\text{Ca}^{2+}$  activity has certain advantages over the others, each also suffers from drawbacks. With the outstanding properties described above, we believe that Quest Rhod-4™ calcium detection reagents and Screen Quest™ Rhod-4NW Calcium Assay Kits provide new powerful tools for intracellular calcium analysis and monitoring in a variety of biological systems.

As might have been predicted, the interests of many researchers in  $\text{Ca}^{2+}$  analysis shifted from the cellular level to the subcellular level. It has been found that  $\text{Ca}^{2+}$  is not even distributed throughout the whole cell and that intracellular heterogeneity of  $\text{Ca}^{2+}$  (such as  $\text{Ca}^{2+}$  waves and  $\text{Ca}^{2+}$  sparks) is observed in a variety of cells (e.g., oocyte, heart muscle cell, hepatocyte, and exocrine cell). With the advent of the confocal laser scanning microscope (CLSM) in the 1980s and advanced microplate readers dedicated for intracellular calcium detection (such as FLIPR™, FDSS and NOVOSTAR™) in 2000s, the measurement of intracellular  $\text{Ca}^{2+}$  has accelerated significantly. Confocal laser scanning microscopy and more recently multiphoton microscopy allow the precise spatial and temporal analysis of intracellular  $\text{Ca}^{2+}$  signaling at the subcellular level.

## Product List

**Table 2** Quest Rhod-4™ Product list

Cat. #	Product Name	Unit Size
21120	Quest Rhod-4™, AM	1 mg
21121	Quest Rhod-4™, AM	5 x 50 µg
21122	Quest Rhod-4™, AM	10 x 50 µg
21123	Quest Rhod-4™, AM	20 x 50 µg
21128	Quest Rhod-4™, sodium salt	5 x 50 µg
36330	Screen Quest™ Rhod-4 NW Calcium Assay Kit *Medium Removal*	1 Plate
36331	Screen Quest™ Rhod-4 NW Calcium Assay Kit *Medium Removal*	10 Plates
36332	Screen Quest™ Rhod-4 NW Calcium Assay Kit *Medium Removal*	100 Plates
36333	Screen Quest™ Rhod-4 NW Calcium Assay Kit *1% FBS Growth Medium*	1 Plate
36334	Screen Quest™ Rhod-4 NW Calcium Assay Kit *1% FBS Growth Medium *	10 Plates
36335	Screen Quest™ Rhod-4 NW Calcium Assay Kit *1% FBS Growth Medium*	100 Plates

## References

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3. Su ZL, Li N, Sun YR, Yang J, Wang IM, Jiang SC. (1998) [Monitoring calcium in outer hair cells with confocal microscopy and fluorescence ratios of fluo-3 and fura-red]. *Shi Yan Sheng Wu Xue Bao*, 31, 323.
4. Perez-Terzic C, Stehno-Bittel L, Clapham DE. (1997) Nucleoplasmic and cytoplasmic differences in the fluorescence properties of the calcium indicator Fluo-3. *Cell Calcium*, 21, 275.
5. Tretyn A, Kado RT, Kendrick RE. (1997) Loading and localization of Fluo-3 and Fluo-3/AM calcium indicators in sinapis alba root tissue. *Folia Histochem Cytobiol*, 35, 41.
6. Greimers R, Trebak M, Moutschen M, Jacobs N, Boniver J. (1996) Improved four-color flow cytometry method using fluo-3 and triple immunofluorescence for analysis of intracellular calcium ion ( $[\text{Ca}^{2+}]_i$ ) fluxes among mouse lymph node B- and T-lymphocyte subsets. *Cytometry*, 23, 205.